

SOME MYXOMYCETES FROM ANDHRA PRADESH, INDIA

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Limited literature is available on floristics of Myxomycetes from India, and no report exists from Andhra Pradesh. Twenty Myxomycetes colonizing diversified habitats have been collected from different forest localities of A.P. and the data has been presented. All the 20 Myxomycetes form new additions to Andhra Pradesh.

KEYWORDS: Andhra Pradesh, Forests, Habitats, Myxomycetes

Myxomycetes (myxa = slime; mycetes = mushrooms, fungi), commonly known as true slime molds were considered earlier as fungi only but now raised to the level of phylum in a separate Kingdom Protozoa (Kirk et al. 2008). These non-chlorophyllous, coenocytic, eukaryotic and multinucleate living organisms' posses a free-living state and exhibit phagotrophic nutrition. Saprobic plasmodium shows a shuttle movement of the protoplasm and plasmodium becomes a resting body or sclerotia under poor conditions. There are four stages in the life cycle of a myxomycete namely: 1) three types of uninucleate cells, one of which is flagellate, 2) multinucleate somatic phase known as plasmodium, 3) resting spores consisting of sclerotia, 4) reproduction leads to stationary sporophores containing walled spores that get disbursed by wind or water and arthropods. Mitotic division shows the presence of non-persistent nuclear membrane (centric) that breaks down in prophase while another being acentric with nuclear membrane being intact like in fungi.

Saprobic plasmodium (the mobile phase of protoplasm) sometimes becoming a resting body or sclerotia under poor conditions. Spores of myxamoebae exist as microcysts and sporangia are either sessile or stalked, often bright coloured. Spores are produced in masses after meiosis and posses persistent or

evanescent peridium. Swarm cells are usually with two anterior flagella and are without cell wall. Spores form myxamoebae directly or after the loss of flagella and sometimes undergo division before copulation. Swarm cells or myxamoebae undergo copulation followed by karyogamy resulting in diploid zygote formation. Zygote on germination gives rise to young plasmodium which becomes mature plasmodium or sclerotium. Sclerotia also get converted into plasmodia which on germination produce sporangia. Meiosis takes place before the mature sporangia are formed. Mature sporangia release spores which on germination release myxamoebae or swarm cells.

Earlier some researchers have reported myxomycetes from India (Lister 1925, Lodhi 1934, Agnihotrudu 1954, 1968, Butler and Bisby 1960, Singh and Pushpavathy 1966, Indira 1971, 1975, Thind 1977, Dhillon and Bermekamp 1978, Bilgrami *et al.* 1979, 1991, Lakhanpal 1983, Nanir 1985, Sarbhoy *et al.* 1975, 1996, Jamaluddin *et al.* 2004, Ranade *et al.* 2012, Tembhurne and Nanir 2011, Tembhurne *et al.* 2012). However, the floristics of myxomycetes has not been explored on par with fungi and algae in India and in particular from Andhra Pradesh. Hence, an attempt has been made to explore the myxomycetes associated with diversified habitats of some forest localities in Andhra Pradesh, India, and the data has been presented.

MATERIALSAND METHODS

Collections were made during July to December (2009-2010). About 120 samples were collected from the forests of Narsapur, Mulugu, Khammam, Araku valley, Ahobilam, Tirumala hills, Vikarabad, Nallamalai and Achampet areas of Andhra Pradesh. Slime molds were collected from dead leaves, twigs, litter, bark, wood logs, etc. Meticulous care was taken during transportation. The collected the Herabrium of Osmania University under OUMH accessions.

RESULTS AND DISCUSSION

Myxomycetes are a cosmopolitian group and are ubiquitous. These are found colonizing litter, bark, wood logs, decaying wood, dead leaves, culms, well manicured lawns, flower beds, humid soils, earthworm casts, dung, dead branches attached to living trees and other substrates. Myxomycetes are distributed in tropics, sub-tropics, temperate forests and also in deserts. Temperature, moisture, rainfall and

Sl. No.	Myxomycete member	Place of collection	Source	Accession No.
1.	Arcyria cinerea (Bull.) Pers.	Pascal forest	Dead wood	OUMH 13
2.	Arcyria nutana (Bull.) Grev.	Araku	Dead wood	OUMH 14
3.	Badhamia papaveracea Berk. & Ravenel	Araku	Wood log	OUMH 16
4.	Ceratiomyxa fruticulosa (Mull.) Macbr.	Bhadrachalam	Dead wood	OUMH 09
5.	Comatricha irregularis Rex	Tirumala hills	Dead wood	OUMH 20
6.	Comatricha laxa Rostaf	Bhadrachalam	Dead wood	OUMH 21
7.	Comatricha typhoides (Bull.) Rostr.	Srisailam forest	Litter	OUMH 22
8.	Cribraria piriformis Schrad	Araku	Dead wood	OUMH 11
9.	Dictydium cancellatum (Batsch) T. Macbr.	Tirumala hills	Dead wood	OUMH 12
10.	Diderma donkii NannBremek.	Vikarabad forest	Dead wood	OUMH 27
11.	Diderma rugosum (Rex) T. Macbr.	Botanical garden, O.U.	Dried leaves, litter	OUMH 19
12.	Lycogala epidendruum (L.) Fries	Bhadrachalam	Dead wood	OUMH 10
13.	Metatrichia resparium Batsch	Vikarabad	Litter	OUMH 15
14.	Physarum echinosporum Lister	Kothagudem	Dead twigs	OUMH 17
15.	Physarum nucleatum Rex	Forest near Ramappa Temple	Unidenti-fied wood	OUMH 18
16.	Stemonitis fusca Roth.	Kothangudem	Dead wood	OUMH 24
17.	Stemonitis oxosora (Bull.)Macbr.	Narsapur forest	Fallen leaves	OUMH 23
18.	Stemonitis pallida Wingate	Vikarabad forest	Dead wood log	OUMH 28
19.	Stemonitis smithii Macbr.	Bhadrachalam	Bark	OUMH 25
20.	Stemonitis splendens Rost.	Ahobilam forest	Dead leaves	OUMH 26

 Table 1. Myxomycetes collected from some forest localities of Andhra Pradesh

samples were preserved in small plastic vials of 2 ½ x 4 cm and were placed vertically in cardboard boxes. Later samples were sun dried and on return were treated with potassium chlorate and calcium chlorate for dehydration by keeping them in desiccators (Davis 1965). For observing calcareous members, Hoyer's medium (Distilled water - 50ml, Arabic gum -30g, chloral hydrate - 200g, glycerin - 20g) was used. Non-calcareous specimens were observed in glycerin mixed with water (40:20). The examined materials have been deposited in other factors are known to influence the periodic occurrence and seasonal variation of species. Agnihotrudu (1954), Indira (1971, 1975), Thind (1977) and Lakhanpal and Mukerji (1981) have made significant contributions to the understanding of myxomycetes from India.

From the Table 1 it is evident that 20 Myxomycetes have been collected from different localities. Among all the substrates, it is noticed that dead wood has been colonized by maximum number of Myxomycetes. It is

interesting to observe that all the twenty Myxomycetes reported form new additions to Myxomycete diversity of Andhra Pradesh, India.

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